

# The effect of variations on protein stability: problems and issues

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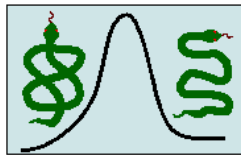


# Outline

1. Dataset influence (*intrinsic to the data*)
2. Biases in the  $\Delta\Delta G$  predictions (*dependent on the method design*)
3. Proper evaluation of the performance (*human behaviour*)

# Datasets

**ProTherm** is a collection of numerical data of thermodynamic parameters including *Gibbs free energy change, enthalpy change, heat capacity change, transition temperature* etc. for wild type and mutant proteins



## ProTherm

### Thermodynamic Database for Proteins and Mutants

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<a href="#">Entry</a>	<input type="text"/>	-	<a href="#">PDB Code</a>	<input type="text"/>	<input type="button" value="Start"/>	<input type="button" value="Clear"/>				
<a href="#">Protein</a>	<input type="text"/>		<a href="#">Source</a>	<input type="text"/>						
<a href="#">Mol-weight</a>	<input type="text"/>	To	<input type="text"/>							
<a href="#">Mutation</a>	<input type="text"/>	To	<input type="text"/>	<input type="checkbox"/> Single	<input type="checkbox"/> Double	<input type="checkbox"/> Multiple	<input type="checkbox"/> Wild Type			
<a href="#">Sec.Structure</a>	<input type="checkbox"/> Helix	<input type="checkbox"/> Sheet	<input type="checkbox"/> Turn	<input type="checkbox"/> Coil						
<a href="#">Accessibility</a>	<input checked="" type="radio"/> Any	<input type="radio"/> Buried	<input type="radio"/> Partially Buried	<input type="radio"/> Exposed	<input type="radio"/> ASA	<input type="text"/>	To	<input type="text"/>	%	<input type="button" value="v"/>
<a href="#">Measure</a>	<input type="checkbox"/> Absorbance	<input type="checkbox"/> CD	<input type="checkbox"/> DSC	<input type="checkbox"/> Fluorescence	<input type="checkbox"/> NMR	<input type="checkbox"/> Others				
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<a href="#">pH</a>	<input type="text"/>	To	<input type="text"/>							

# ProTherm: Thermodynamic Database for Proteins and Mutants

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```
<type 'exceptions.KeyError'>
```

Python 2.7.11: /usr/bin/python2.7  
Fri Feb 17 23:00:09 2017

A problem occurred in a Python script. Here is the sequence of function calls leading up to the error, in the order they occurred.

```
/var/www/cgi-bin/ProTherm/ProTherm.py in ()
 386     df3 = df1.loc[ (df1['muta'].str.contains(muta1,case = False,na=False))& (df1['Type_mutation'].astype(int)== 1)]
 387     else:
=> 388     df3 = df1.loc[pd.to_numeric(df1['Type_mutation'].str.extract('(\d+)')) == 1 & ~df1['muta'].str.contains('wild',case = Fa
 389 elif mutation_type == 'Double':
 390     print"here "
df3 = Empty DataFrame Columns: [] Index: [], df1 = NO. PROTEIN ...2143,2144,2145,21... [25823 rows x 48 columns], df1.loc = <pandas.core.indexing._LocIndexer object>, pd =
from 'usr/local/lib/python2.7/site-packages/pandas/__init__.pyc', pd.to_numeric = <function to_numeric>, .str undefined, case undefined, builtin False = False, na undefined
/usr/local/lib/python2.7/site-packages/pandas/core/frame.py in __getitem__(self= NO. PROTEIN ...2143,2144,2145,21... [25823 rows x 48
columns], key='Type_mutation')
 1962     return self._getitem_multilevel(key)
 1963     else:
=> 1964     return self._getitem_column(key)
 1965
 1966     def _getitem_column(self, key):
self = NO. PROTEIN ...2143,2144,2145,21... [25823 rows x 48 columns], self._getitem_column = <bound method DataFrame._getitem_column of ...143,2144,2145,21... [25823
rows x 48 columns]>, key = 'Type_mutation'
/usr/local/lib/python2.7/site-packages/pandas/core/frame.py in _getitem_column(self= NO. PROTEIN ...2143,2144,2145,21... [25823 rows x 48
```

- 1. Dataset influence (*intrinsic to the data*)**
2. Biases in the  $\Delta\Delta G$  predictions (*dependent on the method design*)
3. Proper evaluation of the performance (*human behaviour*)



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May the *available* experimental measures  
affect the prediction performance?



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# Estimation of a predictor upper bound



# An experimental measure of $\Delta\Delta G$ depends on several factors

- $\Delta\Delta G = f(\text{pH}, T, \text{salts}, C_i, C_j, \dots)$
- In many cases we usually talk only of  $\Delta\Delta G$  which is an average:  
$$\Delta\Delta G = \sum_i \sum_j \sum_k \dots \sum_n \Delta\Delta G(i, j, k, \dots, n)$$
- Sometimes we consider the dependence of pH and T:  
$$\Delta\Delta G = f(\text{pH}, T)$$

# Problem when we compare different measures of the same variation

## Examples

1. In Keeler et al. (2009) the variation H180A in the human prolactin (pdb code 2Q98) measured at  $T=25^{\circ}\text{C}$ , but different pH,  
 $\Delta\Delta G = 1.39 \text{ kcal/mol}$  at pH=5.8;  
 $\Delta\Delta G = -0.04$  at pH=7.8
2. In Gribenko and Makhatadze 2007, the variation E3R in protein 1CSP, 6 different  $\Delta\Delta G$  values ranging from 1.4 kcal/mol to 2.4 kcal/mol were measured at the same temperature ( $55^{\circ}\text{C}$ ) and pH (7.5) as function of different salt concentrations.
3. In Ferguson and Shaw (2002) the variant L3S of the calcium-binding protein S100B (1UWO) measured in two different starting conditions and techniques, but at the same temperature ( $25^{\circ}\text{C}$ ) and pH (7.2) yielded two  
 $\Delta\Delta G = 1.91 \text{ kcal/mol}$  and  
 $\Delta\Delta G = -2.77 \text{ kcal/mol}$

Given the dataset ( $\sigma_{DB}$ ) and the measure uncertainty ( $\sigma$ ) is there an upper bound to the prediction performance?

# Theoretical of estimation of an upper bound: a “*Gedankenexperiment*”

Given N protein variations, we may think to perform a set of N pairs of experiments  $(\{x_i\}, \{y_i\})$ , two for each variation.

Then we use one set of  $\Delta\Delta G$  measures as “predictor” and the other as a set of experimental measures.

The idea is that, given the experimental condition, the best possible predictor is another set of experimental data (considering the experimental uncertainty)

# Theoretical estimation

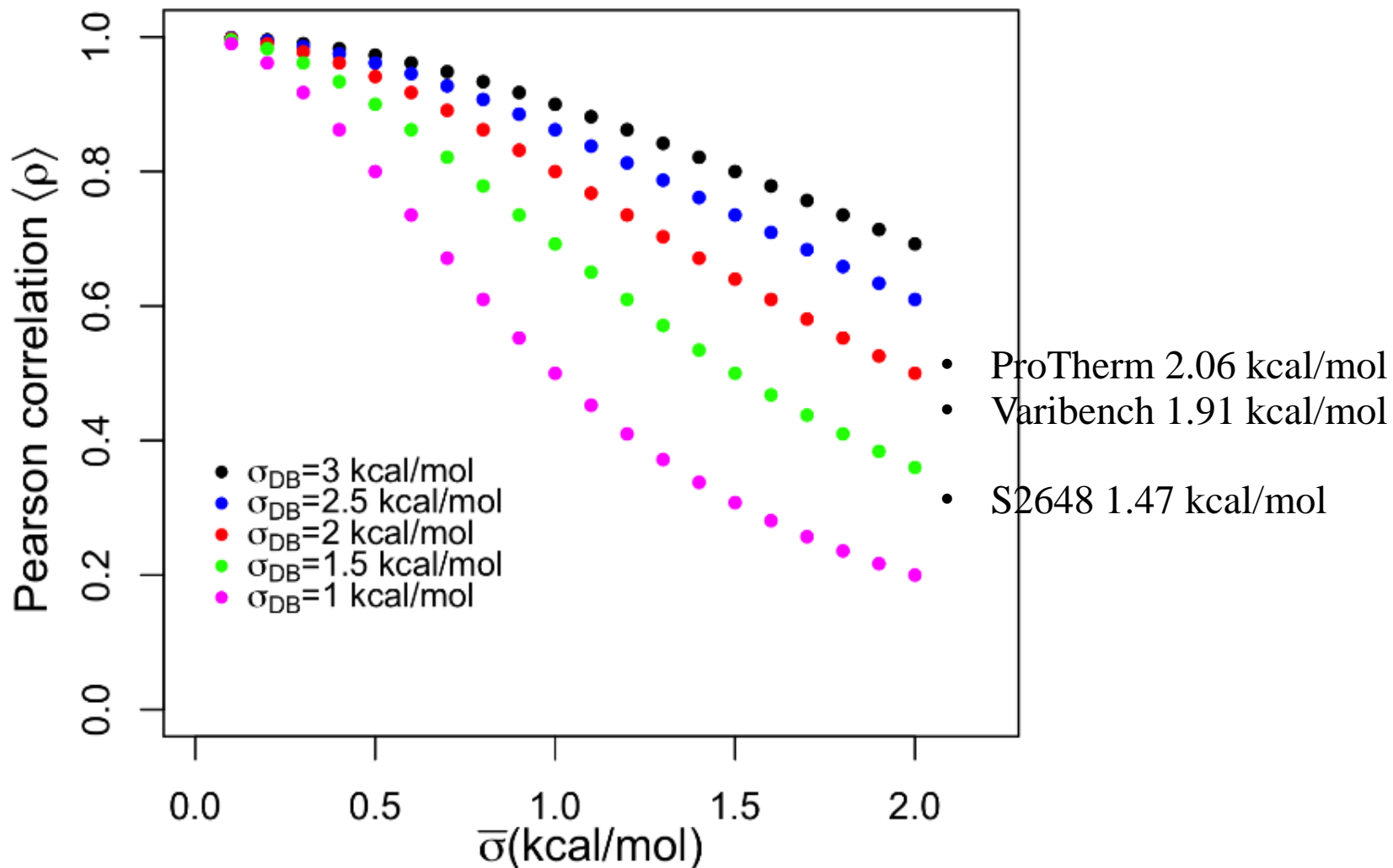
The Pearson's correlation:

$$\langle \rho \rangle \approx \frac{\langle \sigma_{xy} \rangle}{\langle \sigma_x^2 \rangle} \frac{\sigma_{DB}^2}{\overline{\sigma^2} + \sigma_{DB}^2} = \frac{1}{1 + \left( \frac{\overline{\sigma^2}}{\sigma_{DB}^2} \right)}$$

The upper bound of the Coefficient of determination ( $R^2$ ) is even lower than the Pearson with

$$R_{ub}^2 = \langle R^2 \rangle = 1 - \langle S_e \rangle / \langle St \rangle \approx \frac{\sigma_{DB}^2 - \bar{\sigma}^2}{\sigma_{DB}^2 + \bar{\sigma}^2} = \frac{1 - \bar{\sigma}^2 / \sigma_{DB}^2}{1 + \bar{\sigma}^2 / \sigma_{DB}^2}$$

Expected Pearson correlation  $\langle \rho \rangle$  vs. data average uncertainty ( $\bar{\sigma}$ ) for different values of dataset standard deviation  $\sigma_{DB}$



# Experimental Datasets

From

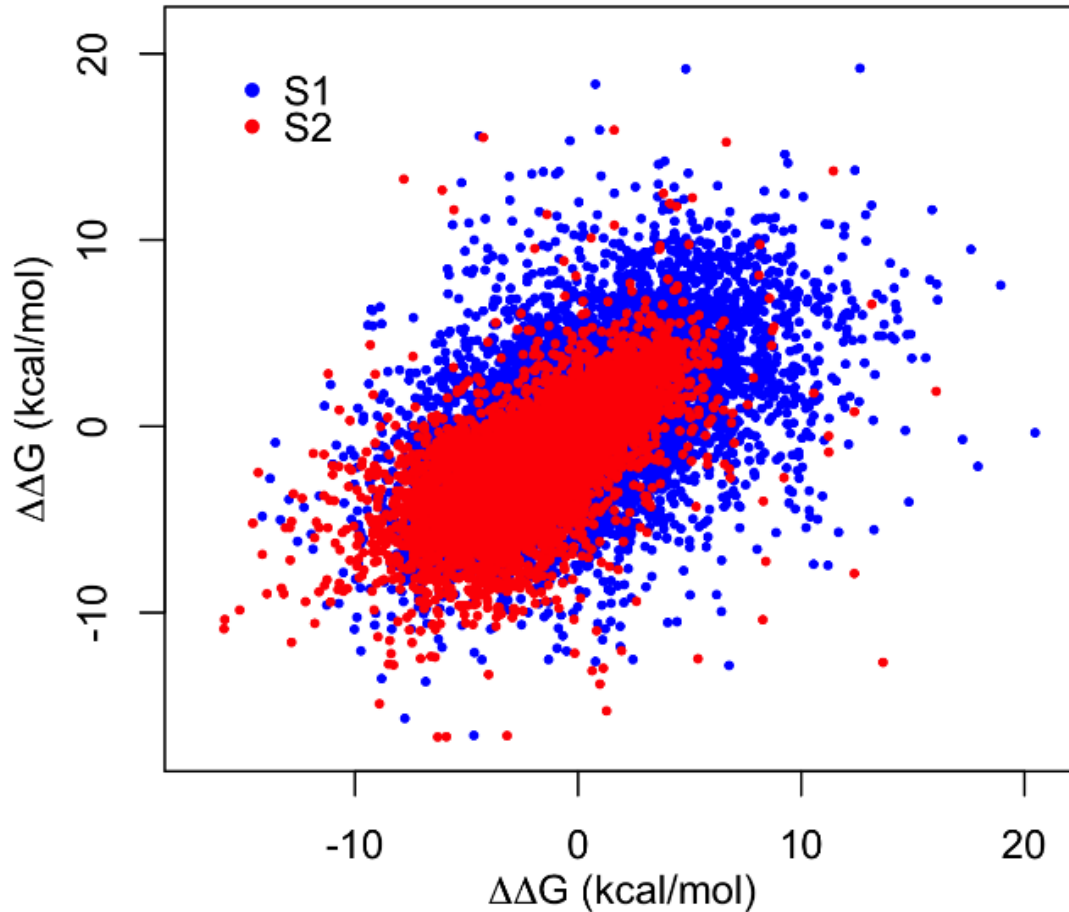
$$\langle \rho \rangle = \frac{1}{1 + \left( \frac{\bar{\sigma}^2}{\sigma_{DB}^2} \right)}$$

and using the experimental data we have

- **S1**: Theoretical estimation with  $\bar{\sigma} = 1.04$  and  $\sigma_{DB}=1.72$   
=> R= **0.73**
- **S2**: Theoretical estimation with  $\bar{\sigma} = 0.72$  and  $\sigma_{DB}=1.57$   
=> R= **0.83**



# Simulation with the experimental Datasets



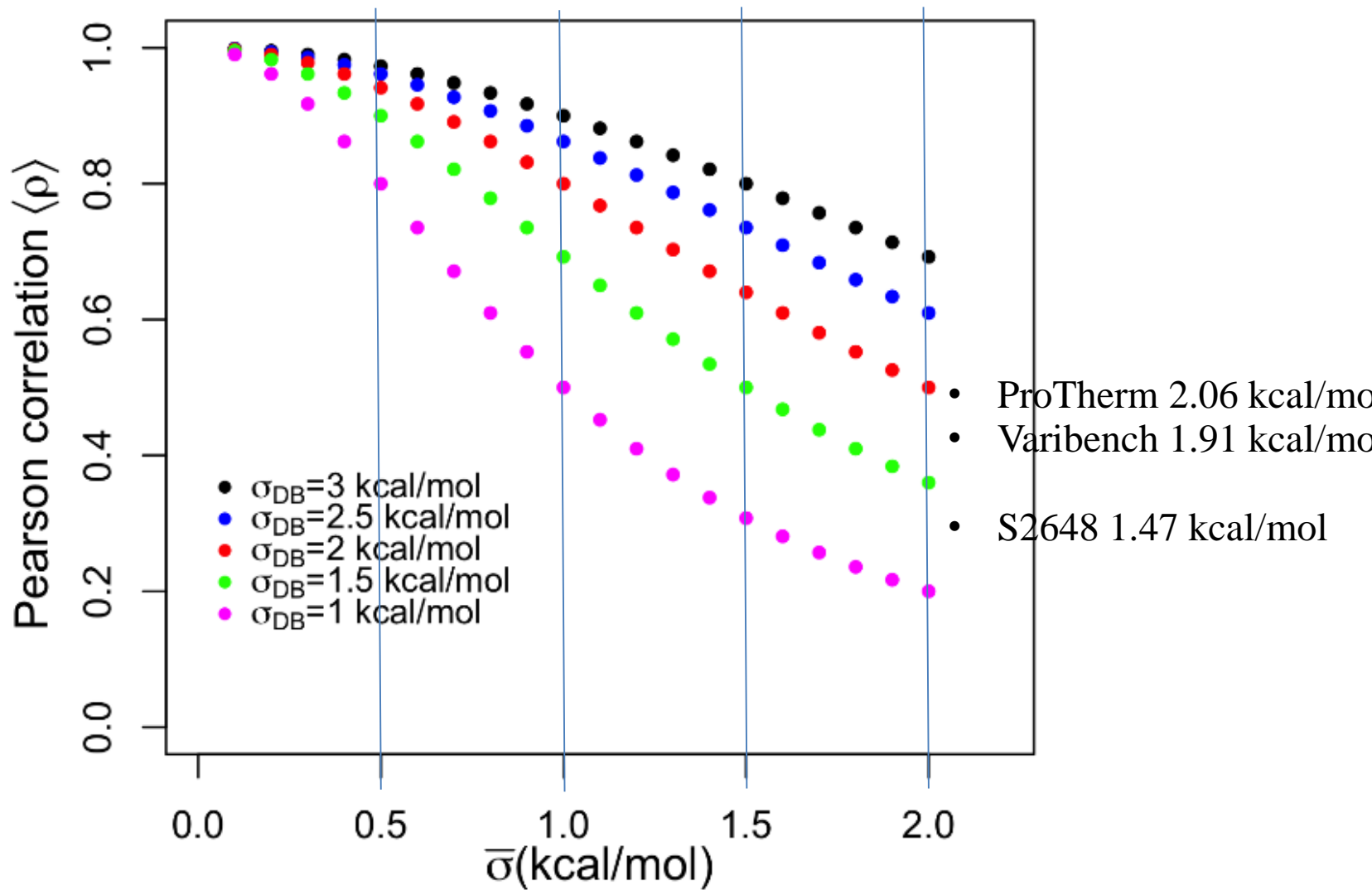
**Scatterplot of two randomly generated observations for a given variation.**

After 100 runs the Pearson correlation are

S1 ->  $0.74 \pm 0.02$

S2 ->  $0.84 \pm 0.02$

# Multiple Mutations ?



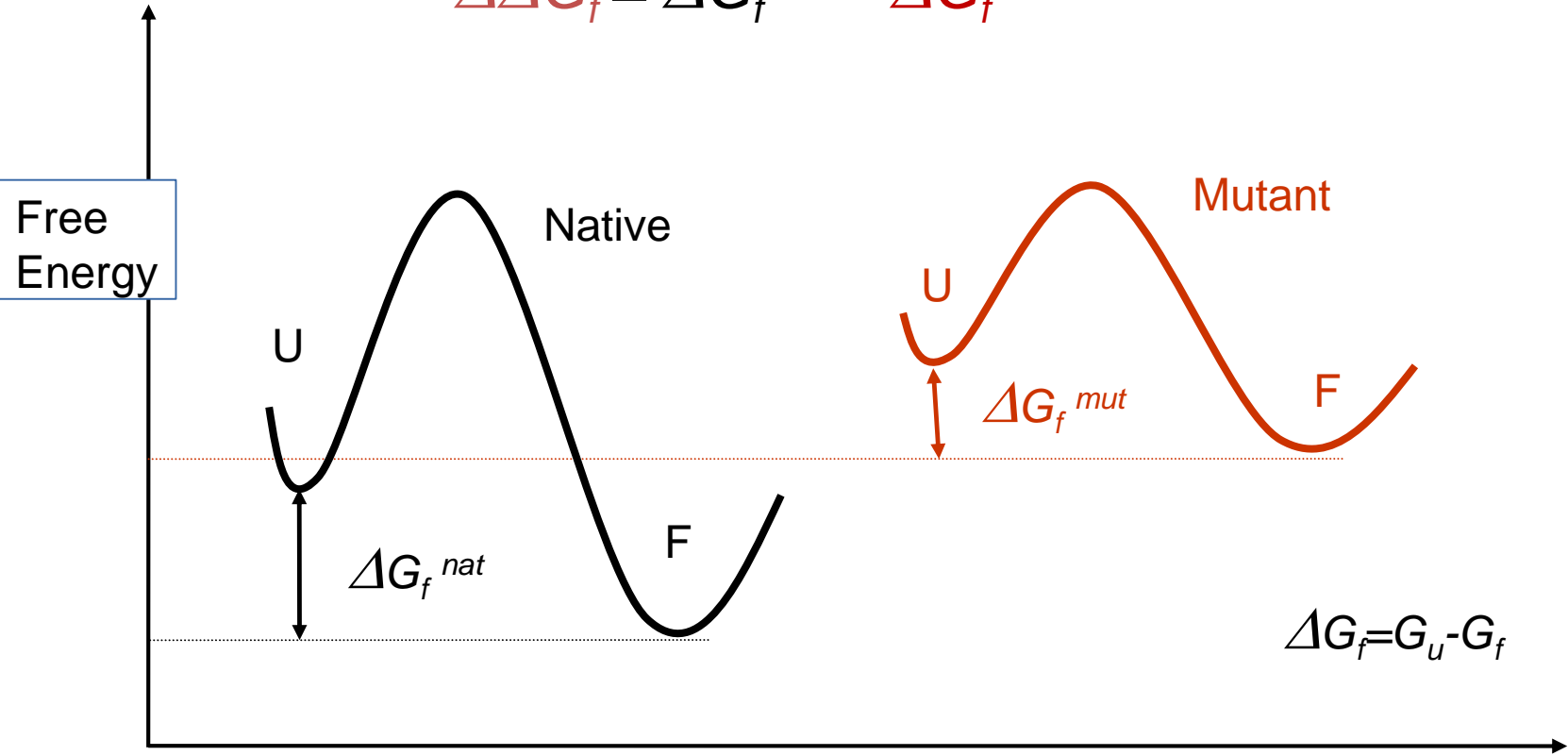
- Comparison among methods on different datasets
- Performance of a method on different datasets
- Evaluate method over-fitting
- Effect on multiple mutations

1. Dataset influence (*intrinsic to the data*)
2. **Biases in the  $\Delta\Delta G$  predictions (*dependent on the method design*)**
3. Proper evaluation of the performance (*human behaviour*)

# Biases in $\Delta\Delta G$ predictions

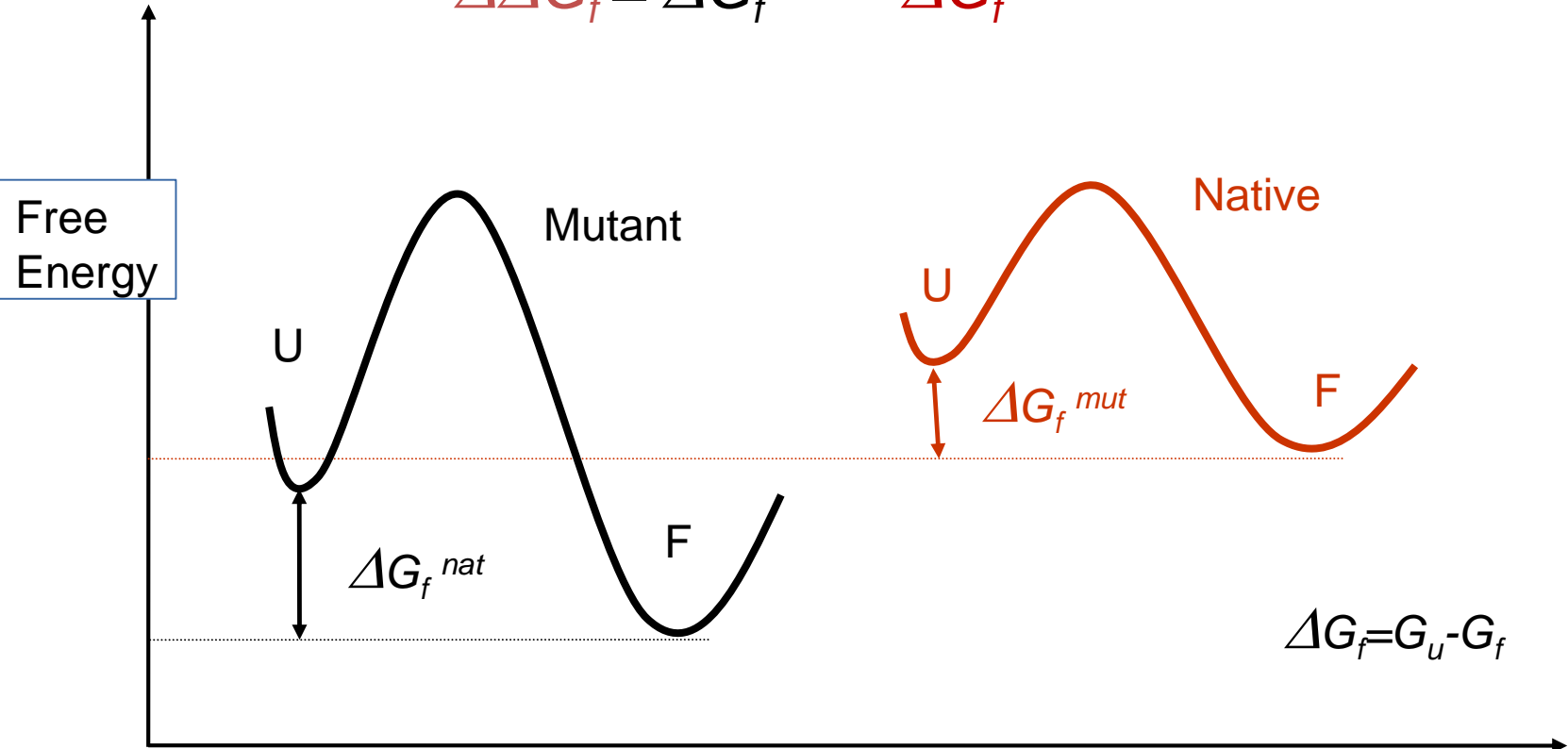
If we change Alanine 35 with a Leucine,  
is the protein stability *Increased or Decreased*?

$$\Delta\Delta G_f = \Delta G_f^{nat} - \Delta G_f^{mut}$$



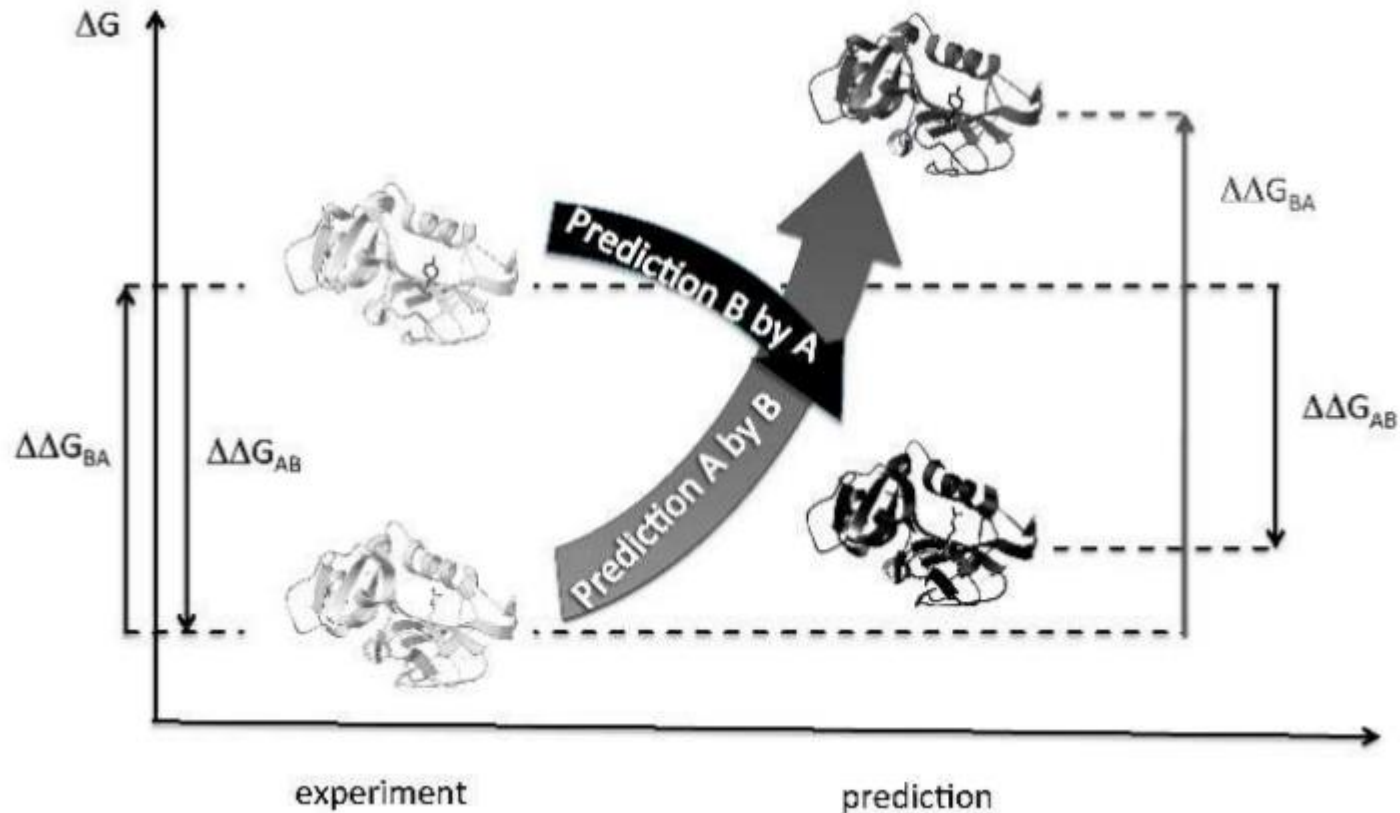
If we change Alanine 35 with a Leucine,  
is the protein stability *Increased or Decreased*?

$$\Delta\Delta G_f = \Delta G_f^{mut} - \Delta G_f^{nat}$$



Protein A = Protein B with variation Y25R  
Protein B = Protein A with variation R25Y

$$\Delta\Delta G_{AB} = -\Delta\Delta G_{BA}$$





# Biases in $\Delta\Delta G$ predictions

Structural Bioinformatics

## Quantification of biases in predictions of protein stability changes upon mutations

Fabrizio Pucci<sup>1</sup>, Katrien Bernaerts<sup>1,2</sup>, Jean Marc Kwasigroch<sup>1</sup> and Marianne Rooman<sup>1</sup>

<sup>1</sup>Department of BioModeling, BioInformatics & BioProcesses, Université Libre de Bruxelles, Roosevelt Ave. 50, 1050 Brussels, Belgium

<sup>2</sup>Biobased Materials, Faculty of Humanities and Sciences, Maastricht University, P.O. Box 616, 6200 MD Maastricht, The Netherlands

## Self-consistency test reveals systematic bias in programs for prediction change of stability upon mutation

Dinara R. Usmanova<sup>1</sup>, Natalya S. Bogatyreva<sup>2,3,4</sup>, Joan Ariño Bernad<sup>5</sup>, Aleksandra A. Eremina<sup>6</sup>, Anastasiya A. Gorshkova<sup>7</sup>, German M. Kanevskiy<sup>8</sup>, Lyubov R. Lonishin<sup>9</sup>, Alexander V. Meister<sup>10</sup>, Alisa G. Yakupova<sup>7</sup>, Fyodor A. Kondrashov<sup>11</sup>, and Dmitry N. Ivankov<sup>4,11,\*</sup>

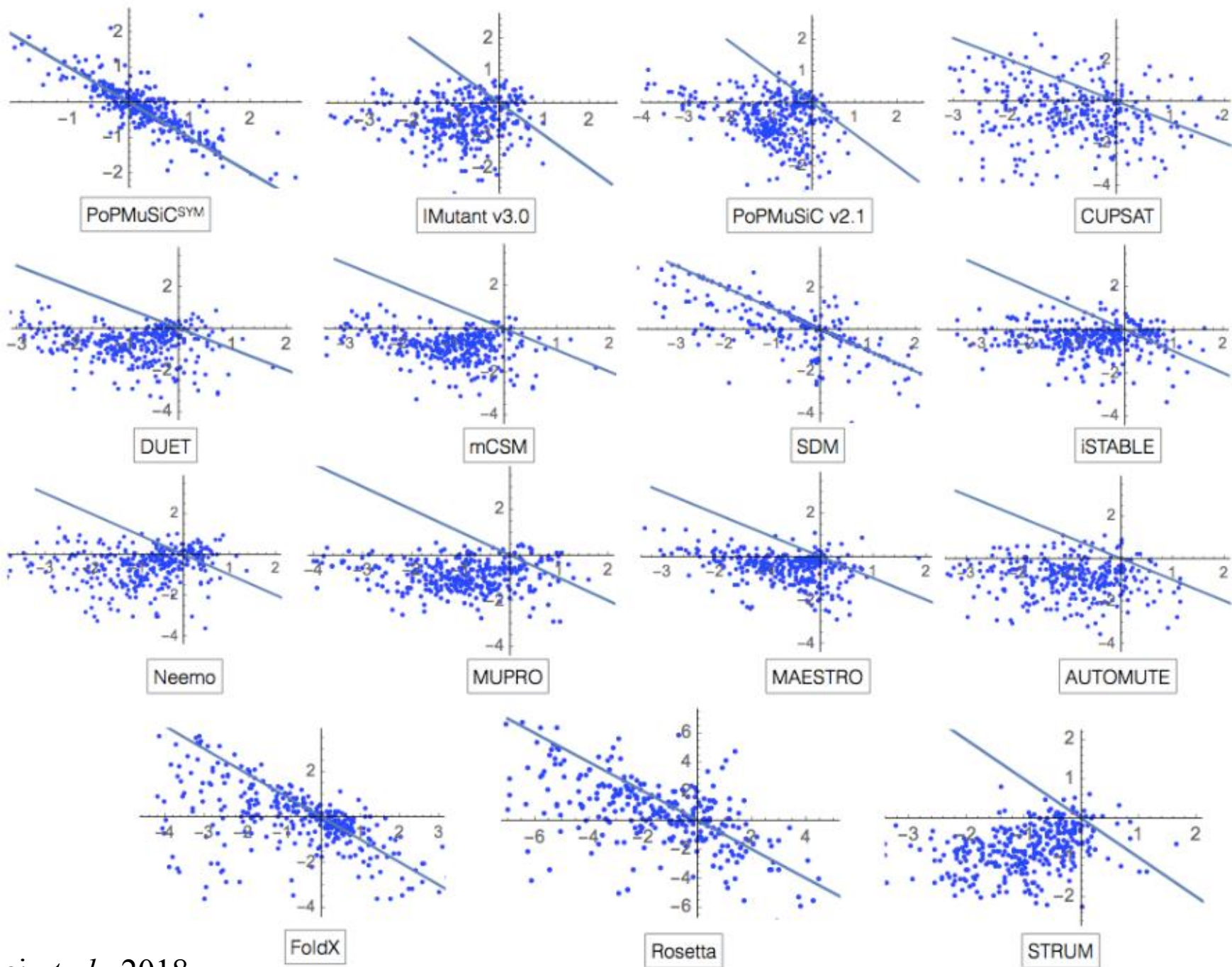
# Biases in $\Delta\Delta G$ predictions

Structural Bioinformatics

## Quantification of biases in predictions of protein stability changes upon mutations

Fabrizio Pucci<sup>1</sup>, Katrien Bernaerts<sup>1,2</sup>, Jean Marc Kwasigroch<sup>1</sup> and Marianne Rooman<sup>1</sup>

- The **Ssym** dataset is a manually curated selection of variations from the ProTherm database.
- It contains mutations with experimental  $\Delta\Delta G$  values for which the 3D structures of both the wild-type and variant proteins were solved by X-ray crystallography.
- **Ssym** consists of 684 variations, 342 are direct (reported in the literature) and 342 are obtained by anti-symmetry, and associated to the variant PDB structure



# Quantification of biases in predictions of protein stability changes upon mutations

Fabrizio Pucci<sup>1</sup>, Katrien Bernaerts<sup>1,2</sup>, Jean Marc Kwasigroch<sup>1</sup> and Marianne Rooman<sup>1</sup>

Method	$\sigma_{\text{dir}}$	$r_{\text{dir}}$	$\sigma_{\text{inv}}$	$r_{\text{inv}}$	$r_{\text{dir-inv}}$	$\langle\delta\rangle$
PoPMuSiC <sup>sym</sup>	1.58	0.48	<b>1.62</b>	<b>0.48</b>	<b>-0.77</b>	<b>0.03</b>
MAESTRO	1.36	0.52	2.09	0.32	-0.34	-0.58
FoldX	1.56	0.63	2.13	0.39	-0.38	-0.47
PoPMuSiC v2.1	1.21	0.63	2.18	0.25	-0.29	-0.71
SDM	1.74	0.51	2.28	0.32	-0.75	-0.32
iSTABLE	1.10	0.72	2.28	-0.08	-0.05	-0.60
I-Mutant v3.0	1.23	0.62	2.32	-0.04	0.02	-0.68
NeEMO	1.08	0.72	2.35	0.02	0.09	-0.60
DUET	1.20	0.63	2.38	0.13	-0.21	-0.84
mCSM	1.23	0.61	2.43	0.14	-0.26	-0.91
MUPRO	<b>0.94</b>	<b>0.79</b>	2.51	0.07	-0.02	-0.97
STRUM	1.05	0.75	2.51	-0.15	0.34	-0.87
Rosetta	2.31	0.69	2.61	0.43	-0.41	-0.69
AUTOMUTE	1.07	0.73	2.61	-0.01	-0.06	-0.99
CUPSAT	1.71	0.39	2.88	0.05	-0.54	-0.72

$$\langle\delta\rangle = \Delta\Delta G_{AB} + \Delta\Delta G_{BA}$$

Table 1. Bias analysis of all the mutations belonging to the dataset  $S^{\text{sym}}$ . The standard deviations  $\sigma_{\text{dir}}$  and  $\sigma_{\text{inv}}$  and the values of  $\langle\delta\rangle$  are in kcal/mol. The methods are ranked according to their performance on the independent test set of inverse mutations, more specifically on the basis of  $\sigma_{\text{inv}}$ .

# Biases in $\Delta\Delta G$ predictions

*Bioinformatics*, YYYY, 0–0

doi: 10.1093/bioinformatics/xxxxx

Advance Access Publication Date: DD Month YYYY

Manuscript Category

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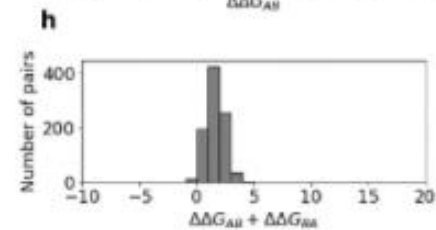
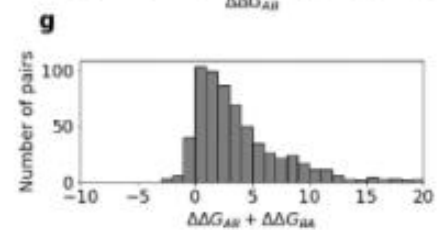
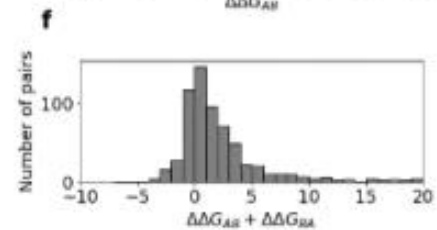
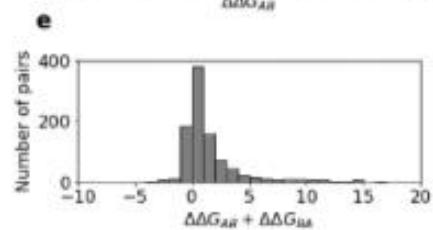
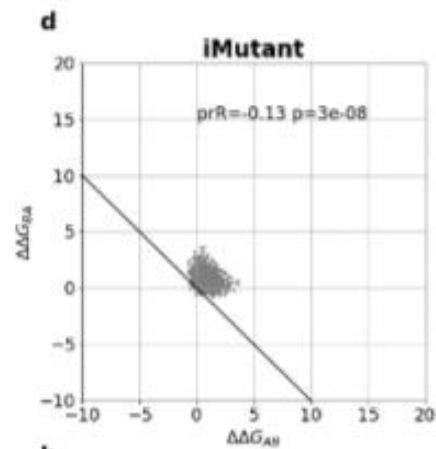
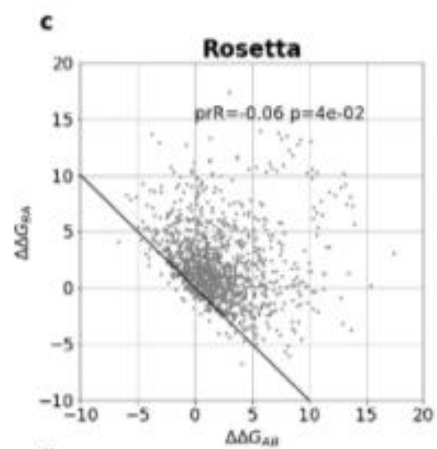
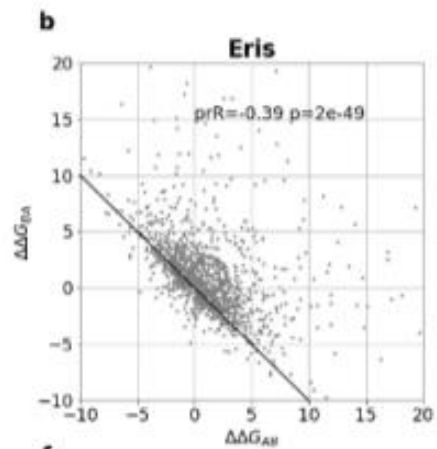
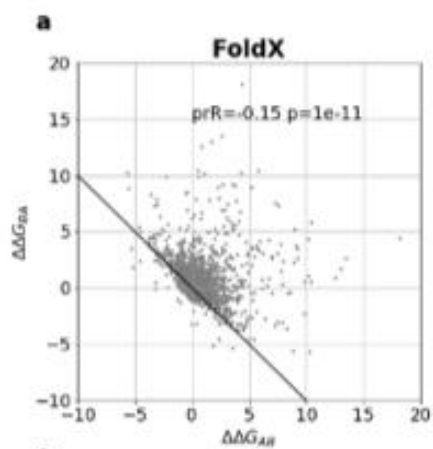
*Subject Section*

## **Self-consistency test reveals systematic bias in programs for prediction change of stability upon mutation**

Dinara R. Usmanova<sup>1</sup>, Natalya S. Bogatyreva<sup>2,3,4</sup>, Joan Ariño Bernad<sup>5</sup>, Aleksandra A. Eremina<sup>6</sup>, Anastasiya A. Gorshkova<sup>7</sup>, German M. Kanevskiy<sup>8</sup>, Lyubov R. Lonishin<sup>9</sup>, Alexander V. Meister<sup>10</sup>, Alisa G. Yakupova<sup>7</sup>, Fyodor A. Kondrashov<sup>11</sup>, and Dmitry N. Ivankov<sup>4,11,\*</sup>

# Biases in $\Delta\Delta G$ predictions

- dataset was built by Usmanova *et al.* 2018, by extracting high-resolution pairs of proteins from the Protein Data Bank (PDB) differing by one to ten amino acids.
- Large datasets, with 1000 pairs of protein structures differing by one residue



## **Self-consistency test reveals systematic bias in programs for prediction change of stability upon mutation**

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**Table 1.** Bias for single substitutions

Program	Bias, kcal/mol	r (p-value)
FoldX	0.74 ± 0.05	-0.15 (10 <sup>-11</sup> )
Eris	1.25 ± 0.11	-0.39 (2·10 <sup>-49</sup> )
Rosetta	2.08 ± 0.12	-0.06 (0.04)
I-Mutant	0.80 ± 0.01	-0.13 (3·10 <sup>-8</sup> )

$$\text{Bias} = (\Delta\Delta G_{AB} + \Delta\Delta G_{BA})/2$$



An important property that a predictor has to fulfil is the “variation” anti-symmetry :

$$\Delta\Delta G_{AB} = - \Delta\Delta G_{BA}$$

# DDGun: DDG Untrained baseline method

A way to implement the predictor anti-symmetry is to provide in input to it only **anti-symmetric features**

# DDGun: DDG Untrained baseline method

Assuming that the profile  $p$  does not change for the mutant and the wild type protein sequence, we can compute some feature scores such as

Evolutionary  
( $B$ =Blosum62)

$$s_{Bl} = \sum_{i=1}^{20} p(a_i)(B(a_i, m) - B(a_i, w))$$

Skolnick ( $P_{Sk}$ )  
Local potential

$$s_{Sk} = \sum_{j=-2}^{j+2} \sum_{j \neq 0} p(a_j)(P_{Sk}(w, a_i) - P_{Sk}(m, a_i))$$

Hydrophobicity ( $K$ )

$$s_{Hp} = p(m)K(m) - p(w)K(w)$$

3D contact potential ( $P_{BV}$ )

$$s_{BV} = \sum_{j \in I} \sum_{i=1}^{20} p(a_{ij})(P_{BV}(w, a_i) - P_{BV}(m, a_i))$$

# DDGun: DDG Untrained baseline method

Anti-symmetry performances of DDGun on the Ssym data set (Pucci et al, 2018, PopMusicSym)

Method	Performances		Anti-symmetry	
	Direct variations Pearson r, RMSE	Inverse variations Pearson r, RMSE	$r_{\text{dir-inv}}$	Bias $\langle \delta \rangle$ (kcal/mol)
DDGun	0.48, 1.47	0.48, 1.50	-0.99	-0.007
DDGun3D	0.56, 1.42	0.53, 1.46	-0.99	-0.02
PopMusicSym	0.48, 1.58	0.48, 1.62	-0.77	0.03
SDM	0.51, 1.74	0.32, 2.28	-0.75	-0.32
Maestro	0.52, 1.36	0.32, 2.09	-0.34	-0.58
FoldX	0.63, 1.56	0.39, 2.13	-0.38,	-0.47

# DDGun: DDG Untrained baseline method

Performances on the 914 multiple site variation from Protherm.

Method	Performances			Anti-symmetry	
	Direct and Inverse Pearson r, RMSE	Direct variations Pearson r, RMSE	Inverse variations Pearson r, RMSE	$r_{\text{dir-inv}}$	Bias $\langle\delta\rangle$ (kcal/mol)
DDGun	0.44, 2.23	0.37, 2.23	0.37, 2.23	-1.00	0.00
DDGun3D	0.45, 2.27	0.39, 2.24	0.38, 2.25	-0.99	-0.007
Maestro	0.30, 2.59	0.55, 1.96	0.08, 3.10	-0.20	-0.92
FoldX	0.44, 3.10	0.41, 2.95	0.33, 3.24	-0.71	-0.21

1. Dataset influence (*intrinsic to the data*)
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# Evaluation problems:

1. many Variations in the same (similar) protein
2. many Variations in the same protein position

*Classical mistake:* random partition of training and testing sets to fit the parameters or train models

Problem of similarity between  
training and testing sets



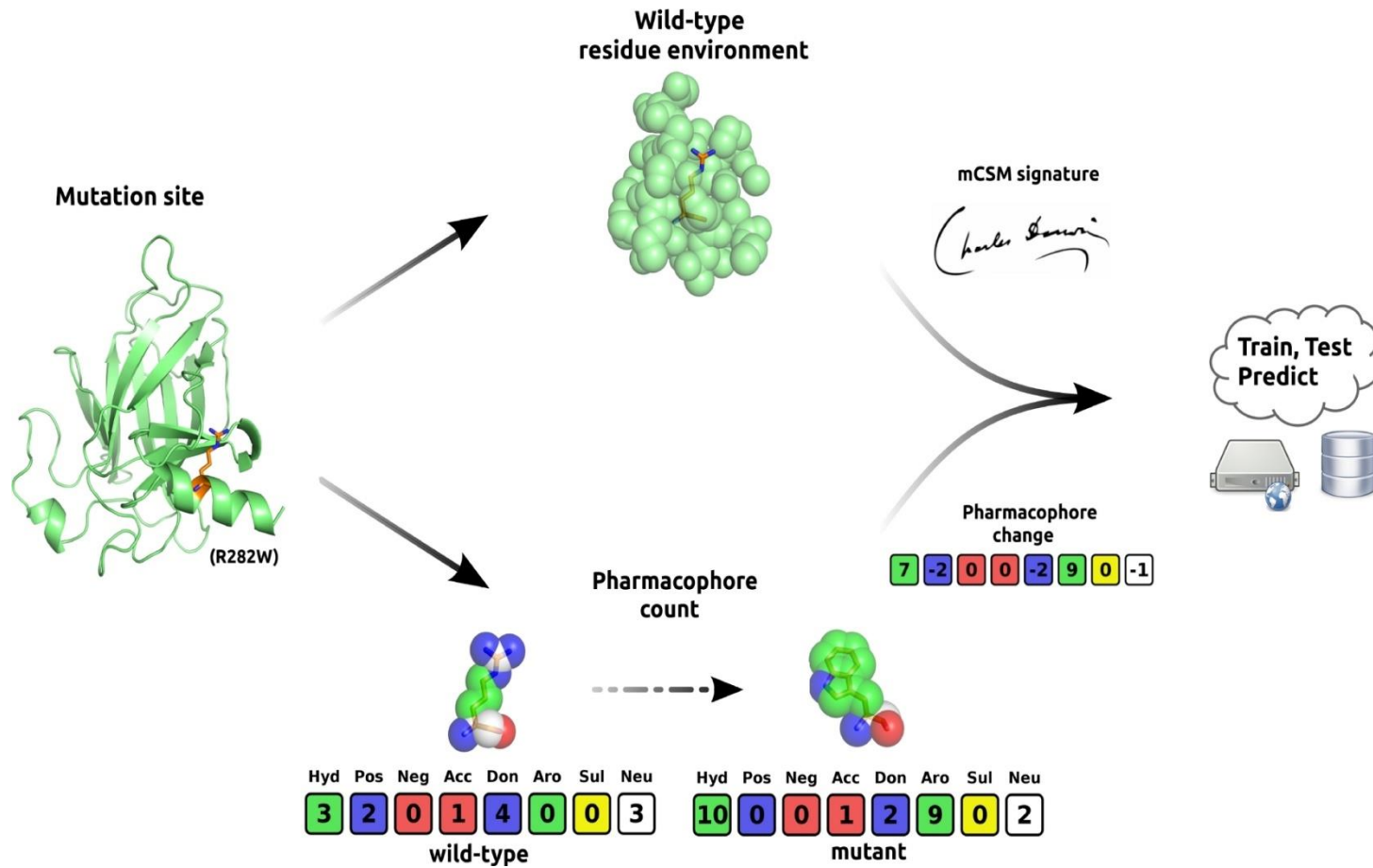
## **Protein stability: a single recorded mutation aids in predicting the effects of other mutations in the same amino acid site**

Gilad Wainreb<sup>1</sup>, Lior Wolf<sup>2,\*</sup>, Haim Ashkenazy<sup>1</sup>, Yves Dehouck<sup>3</sup> and Nir Ben-Tal<sup>1,\*</sup>

<sup>1</sup>Department of Biochemistry and Molecular Biology, George S. Wise Faculty of Life Sciences, <sup>2</sup>The Blavatnik School of Computer Science, Tel-Aviv University, Ramat Aviv 69978, Israel and <sup>3</sup>Bioinformatique génomique et structurale, Université Libre de Bruxelles, Av Fr. Roosevelt 50, CP165/61, 1050 Brussels, Belgium

Associate Editor: Anna Tramontano

# The case of mCSM

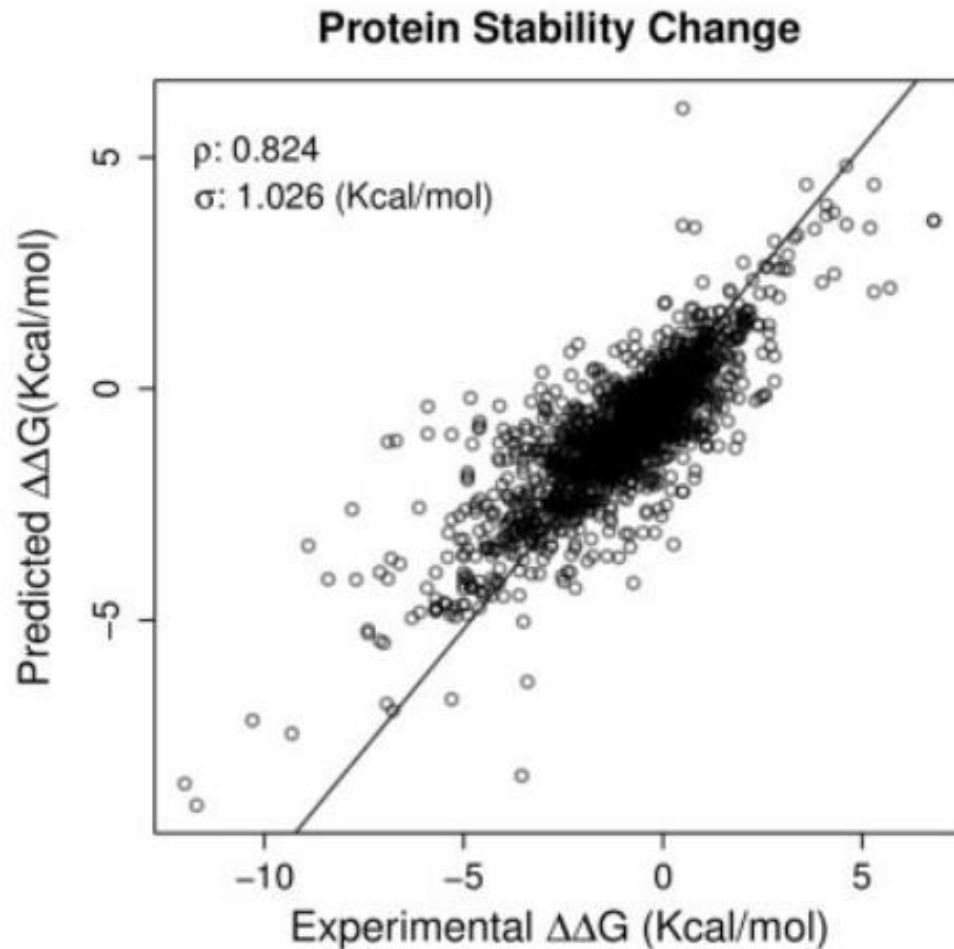


(\*) mCSM: predicting the effect of mutations in proteins using graph-based signatures

Douglas E. V. Pires, David B. Ascher, Tom L. Blundell, 2014

# mCSM

## main paper results



(\*) mCSM: predicting the effect of mutations in proteins using graph-based signatures

Douglas E. V. Pires, David B. Ascher, Tom L. Blundell, 2014

**Fig. 2.** Regression results for mCSM signature pred

# mCSM

**Table 2.** Comparative regression experiments using the S350 data set

Method	Number of predictions	Pearson's coefficient <sup>a</sup>	Standard error(kcal/mol) <sup>a</sup>
Automute	315	0.46/0.45/0.45	1.43/1.46/1.99
Cupsat	346	0.37/0.35/0.50	1.91/1.96/2.14
Dmutant	<b>350</b>	0.48/0.47/0.57	1.81/1.87/2.31
Eris	334	0.35/0.34/0.49	4.12/4.28/3.91
I-Mutant-2.0	346	0.29/0.27/0.27	1.65/1.69/2.39
PoPMuSiC-1.0	<b>350</b>	0.62/0.63/0.70	1.24/1.25/1.66
PoPMuSiC-2.0	<b>350</b>	0.67/0.67/0.71	1.16/1.19/1.67
SDM	<b>350</b>	0.52/0.53/0.63	1.80/1.81/2.11
<b>mCSM</b>	<b>350</b>	<b>0.73/0.74/0.82</b>	<b>1.08/1.10/1.48</b>

*Note:* Results directly obtained from Worth *et al.* (2011). Bold values highlight are the best performing metrics.

<sup>a</sup>The three values given per column correspond, respectively, to the whole validation set of 350 mutants, the 309 mutants for which a prediction was available for all predictors. Finally, in the third column are the results for 87 mutants, a subset of the

main paper  
results

(\*) mCSM:  
predicting the effect  
of mutations in  
proteins using  
graph-based  
signatures

Douglas E. V. Pires,  
David B. Ascher, Tom  
L. Blundell, 2014

# mCSM

## Supplementary material

**Table 9.** Evaluation of predictive performance of mCSM for the S2648 data set in new low-redundancy blind and cross validation schemes. Results are given for data set partitioning in Protein (Prot) and Position (Pos) levels as described in Section 4.2.

Method	Data set	Validation	Pearson's coeff.*	Std. error(Kcal/mol)*
mCSM	S2648	5-fold (Pos)	0.54/0.69	1.23/0.90
mCSM	S2648	5-fold (Prot)	0.51/0.66	1.26/0.94

(\*) mCSM: predicting the effect of mutations in proteins using graph-based signatures

Douglas E. V. Pires, David B. Ascher, Tom L. Blundell, 2014

# mCSM

## Summary:

- Training  $\rightarrow r = 0.82$
- Random split  $\rightarrow r = 0.73$
- CV for positions  $\rightarrow r = 0.54$
- CV for proteins  $\rightarrow r = 0.51$

**(\*) mCSM: predicting the effect of mutations in proteins using graph-based signatures**

Douglas E. V. Pires, David B. Ascher, Tom L. Blundell, 2014

# mCSM

Other examples with meta-predictors



## Computational tools help improve protein stability but with a solubility tradeoff

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Edited by Wolfgang Peti

Tool	MCC	R	Precision	Accuracy	S.E.
			%	%	<i>kcal/mol</i>
EGAD	0.34	0.52	50	74	1.61
FoldX	0.38	0.54	52	78	1.78
Rosetta-ddG	0.32	0.54	46	75	2.34
CUPSAT	0.24	0.55	44	75	1.77
DFire	0.43	0.64	49	76	1.84
Hunter	0.16	0.32	34	68	1.89
MultiMutate	0.19	0.54	32	62	2.34
SDM	0.26	0.46	37	68	1.96
PoPMuSiC	0.33	0.68	59	79	1.32
IMutant3	0.14	0.51	41	75	1.52
MuPro	0.18	0.49	57	78	1.52
Meta-predictor	<b>0.48</b>	<b>0.73</b>	<b>63</b>	<b>82</b>	<b>1.29</b>

- 60% of the proteins are in the training of some predictors
- The Meta-predictor was trained on 50% of randomly selected data and tested on the other 50% (similarity issue)



## **DynaMut: predicting the impact of mutations on protein conformation, flexibility and stability**

**Carlos H.M. Rodrigues<sup>1</sup>, Douglas E.V. Pires<sup>2,\*</sup> and David B. Ascher<sup>1,2,3,\*</sup>**

<sup>1</sup>Department of Biochemistry and Molecular Biology, Bio21 Molecular Science and Biotechnology Institute, University of Melbourne, Australia, <sup>2</sup>Instituto René Rachou, Fundação Oswaldo Cruz, Brazil and <sup>3</sup>Department of Biochemistry, University of Cambridge, UK

- **Dataset S2648**
- Data presented in:
  - training on the data
  - cross-validation with random split
  - random generation on a “blind” set of 351 variations from S2648, and trained on the remainder 2297 variants.
- Based on the output of DUET, SDM2, mCSM that were **TOTALLY** trained on **S2648**

**Please:** *test your model using data (predictors) that have no sequence similarity (trained on proteins similar) to those of your test set!*

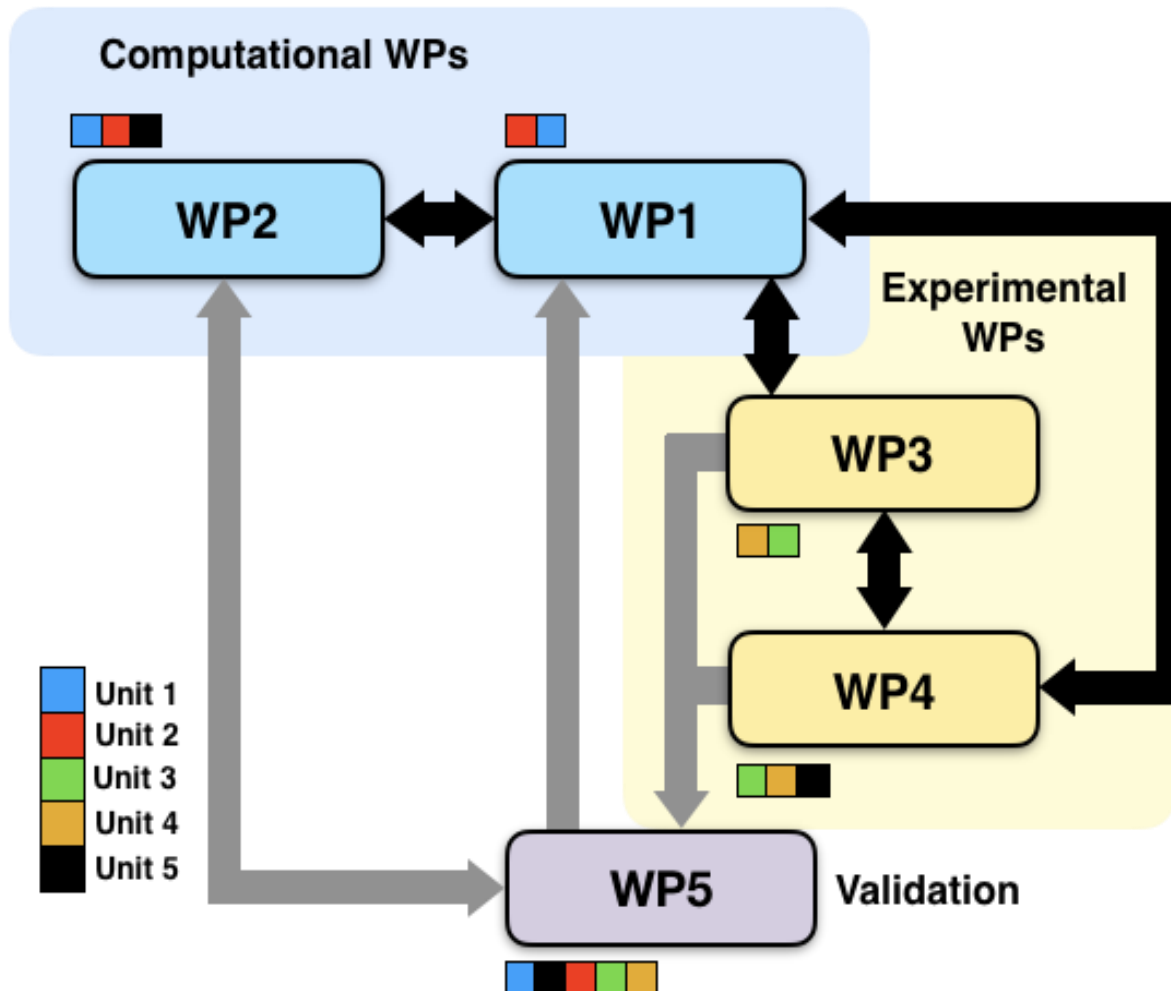
# Contributors



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# Project:



# Project:

	Year I												Year II												Year III													
	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36		
WP1	Database implementation and development																																					
WP2					Developments of the Predictors																																	
WP3	Generation of new experimental data: structural, functional and stability																																					
WP4					Generation of new experimental data: binding affinity variations.																																	
WP5																									Data validation and evaluation of the predictors													
Deliverables WP1					■								■														■											
Deliverables WP2											■								■								■											
Deliverables WP3	■								■								■																					
Deliverables WP4											■								■								■											
Deliverables WP5																									■													
Meetings											■														■													
Seminars											■								■								■											
Dissemination											■														■													
Public Engagement											■								■								■											
Reports											■														■													

# Project:

Definition of the objectives and deliverables in the light of the 38% cut